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9β-LANOSTANE-TYPE TRITERPENE LACTONES FROM THE STEM BARK OF ABIES VEITCHII

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ABSTRACT.—Two new tetracyclic triterpene lactones were isolated, together with two known compounds, 3-oxo-9 β -lanosta-7,24-dien-26,23R-olide [**2**] and 3 β -hydroxy-9 β -lanosta-7,24-dien-26,23R-olide [**3**], from the stem bark of *Abies veitchii*. The structures of the new compounds were established as 3 α -hydroxy-9 β -lanosta-7,24-dien-26,23R-olide [**6**] and 3 α -methoxylanosta-7,9(11),24-trien-26,23R-olide [**7**], respectively, on the basis of chemical and spectral evidence.

Abies veitchii Lindl. (Pinaceae) (Japanese name: Shirabiso) is a tall evergreen tree growing deep in the mountains from central to northern Japan (1). Our previous studies on the chemical constituents of Abies species have revealed a tetracyclic triterpene lactone, abieslactone [1] $(3\alpha$ -methoxy-9 β -lanosta-7,24-dien-26,23R-olide) (2,3), from the bark and leaves of Abies mariesii Mast., Abies amabilis (Dougl.) Forbes, and Abies procera Rehd., a mixture of five *n*-alkyl ferulates bearing alkyl moieties from C-22 to C-26, a mixture of campesterol and β -sitosterol, 3-oxo-9 β -lanosta-7,24-dien-26,23R-olide [2] (4), 3β -hydroxy- 9β -lanosta-7, 24-dien-26, 23*R*-olide [3], and 27-hydroxy-3-oxo-9β-lanosta-7,24-dien-26,23R-olide [4] (5) from the bark of Abies firma Sieb. et Zucc., and abieslactone [1] and veitchiolide [5] $\{7\beta$ -hydroxy- 3α -methoxylanosta-9(11),24dien-26,23R-olide] from the bark of A. veitchii (6). The fact that the bark of A. firma contained substantial 2 and lacked abieslactone [1], which is the most abundant triterpenoiod in the bark of both A. mariesii and A. veitchii, drew our chemotaxonomic interest in this species. During the course of our work, five groups of investigators have reported the isolation and the structure elucidation of several highly oxygenated lanostanes, 9B-lanostanes, cycloartanes, and some of their analogues from the bark and needles of Abies alba Mill. (7,8), the bark of A. grandis (9), the seeds of both A. mariesii and A. firma (10, 11), and the bark and needles of Abies sibilica (12, 13).

Recently, we reported that methyl esters **1a** and **2a** of two γ -keto acids and a trisnor- α -hydroxy acid **1b**, derived from compounds **1** or **2**(2), strongly inhibited both (a) incorporation of ³²P into phospholipids of HeLa cells cultured in the medium containing 12-0-tetradecanoylphorbol-13-acetate (TPA) and inorganic ³²P and (b) the promoting action of TPA on skin tumor formation in mice initiated with 7, 12-dimethyl-



benz[a]anthracene (14), although compounds 1 and 2 themselves showed no remarkable activity in the same in vitro assay.

In the search for stronger and more effective antitumor-promoting agents from plant sources, we re-examined the neutral Et_2O extract of the stem bark of *A. veitchii* and isolated two unknown triterpenoid lactones **6** and **7**, together with two known triterpene constituents **2** and **3**. This paper deals with the structure elucidation of compounds **6** and **7**.



RESULTS AND DISCUSSION

The known triterpenoids were identified by direct comparison with samples of 3oxo-9 β -lanosta-7,24-dien-26,23*R*-olide [**2**] and 3 β -hydroxy-9 β -lanosta-7,24-dien-26,23*R*-olide [**3**], respectively, isolated from the bark of *A*. firma (4,5).

Compound 6, one of two unknown compounds, gave the molecular formula $C_{30}H_{46}O_3$ ([M]⁺ at m/z 454.3443) and showed positive color on the Liebermann-Burchard test. Its ir, ¹H-nmr (Table 1), and ¹³C-nmr spectra (Table 2) exhibited the presence of five tertiary methyl groups, a secondary methyl group, a secondary carbinolic methine group [δ 3.43 (1H, narrow diffused t, W/2 = 6.8 Hz) and 76.32 (H-C-OH)], a trisubstituted olefin bond [δ 5.55 (1H, H-7), 121.64 (=CH-) and 148.60 (=C<)], along with signals due to the presence of a 4-substituted 2-methyl-2-butenolide ring (closely similar to that in both compounds 2 and 3) at δ 1.91 (3H, t, Me-27), 4.98 (1H, ddd, H-23), 7.00 (1H, H-24), 129.46 (=C<) and 149.72 (=CH-). In the DEPT and eims, compound $\mathbf{6}$ showed the same carbon composition and fragment ion peaks (Scheme 1) closely similar to those of compound 3(5). However, ¹H-nmr chemical shift values for H-3, Me-19, Me-28, and Me-29 in compound 6 showed significant discrepancies of $\Delta \delta + 0.17$, +0.08, +0.12, and -0.09 ppm, respectively, in comparison with those of compound 3, although all the other ¹H signals in both compounds agreed within ± 0.01 ppm. Furthermore, ¹³C-nmr signals of C-1, C-2, C-3, C-4, C-5, C-19, C-28, and C-29 around the A ring in compound 6 showed chemical shift values considerably different from those of compound $\mathbf{3}$, whereas all the other signals of both compounds were in good agreement. Acetylation of compound 6 yielded 6-acetate. In the ¹H-nmr spectrum, compounds 6 and 6-acetate exhibited signals attributable to 3-equatorial methine protons geminal to the hydroxyl group and acetoxy group as narrow diffused triplets (W/2 = 6.8 Hz) at δ 3.43 and 4.65, respectively, while compound 3 showed the corresponding 3-axial methine proton signal as a double doublet (J = 10.2 and 5.5 Hz) at $\delta 3.20$ (5). All these data supported the theory that compound 6 must be the 3α isomer of 3. The 23R configuration of the lactone side chain in compound 6 was estimated from cd measurements in which it gave a negative Cotton effect curve similar to those of compounds 2 and 3(4,5). Definite proof for this assumption was obtained by the following experiment. Chromium trioxide oxidation of 6 in pyridine afforded the keto-lactone identical in all respects with compound 2. Accord-

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of Compounds
Shifts o
Chemical
H-nmr
TABLE 1.

Proton			Compo	pund		
	2	æ	Sª	6	6-acetate	7
Me-18	0.81	0.93	0.69	0.943	0.95	0.59
Me-19	1.00	0.93	1.08	1.01	1.02	1.00
Me-21	1.01	1.00	1.03	1.00	1.00	1.02
	(d, 6.5)	(d, 6.5)	(d, 6.5)	(d, 6.5)	(q, 6.5)	(d, 6.5)
Me-27	1.92	1.91	1.92	1.92	1.91	1.92
	(t, 1.7)	(t, 1.7)	(t, 1.7)	(t, 1.7)	(t, 1.7)	(t, 1.7)
Me-28	1.10	0.87	0.94	0.99	0.88	0.948
Me-29	1.09	1.02	0.89	0.925	0.98	0.934
Me-30	1.03	1.02	0.83	1.03	1.02	0.87
Η-2α,β	2.49	ł	I			
	(dd, 8.5, 6.5)					
Н-3		3.20	2.84	3.43	4.65	2.84
		(dd, 10.0, 5.5)	(t, 2.4)	(dif. t, W/2 6.8)	(dif. t, W/2 6.8)	(dif. t, W/2 6.8)
H-7	5.64	5.56	3.71	5.55	5.55	5.31
-	(dt, 6.5, 3.0)	(dt, 6.5, 3.0)	(td, 11.5, 5.3)	(dt, 6.5, 3.0)	(dt, 6.5, 3.0)	(br. d, 6. l)
H-11			5.31			5.44
		_	(br. d, 6.3)			(t, 4.5)
H-23	4.98	4.97	4.99	4.98	4.97	4.98
	(ddd, 9. 4, 4. 1, 1.7)	(ddd, 9.4, 4.1, 1.7)	(ddd, 9.4, 4.1, 1.7)	(ddd, 9.4, 4.1, 1.7)	(ddd, 9.4, 4.1, 1.7)	(ddd, 9.5, 4.1, 1.7)
H-24	7.01	6.99	7.00	7.00	7.00	7.01
	(quint, 1.7)	(quint, 1.7)	(quint, 1.7)	(quint, 1.7)	(quint, 1.7)	(quint, 1.7)
3α-OMe	Ι		3.31	1		3.29
3α-OAc					2.05	
^a Data for this c	compound are taken fror	n Tanaka and Matsuna,	ga (6).			

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SCHEME 1. Mass spectral fragmentation of compounds 6 and 6-acetate.

ingly, the structure of compound **6** was proved to be 3α -hydroxy-9 β -lanosta-7,24-dien-26,23*R*-olide.

Compound 7, the second new compound, gave the molecular formula $C_{31}H_{46}O_3$ $([M]^+$ at m/z 466.3451) and also was positive in the Liebermann-Burchard test. Its uv and it spectra showed the presence of an α,β -unsaturated γ -lactone ring [λ max 219 nm; ν max 1758 (shoulder) and 1740 cm⁻¹] and a heteroannular diene chromophore (λ max 236, 243.5 and 251.5 nm; ν max 1658, 897 and 815 cm⁻¹) in the molecule. In the ¹H- and ¹³C-nmr spectra, it exhibited signals due to five quaternary methyl groups, a secondary methyl group, a vinylic methyl group deshielded with the lactone carbonyl, a secondary methoxy group attached to the C-3 α position [δ 2.84 (1H, narrow diffused t, H-C-OMe), 3.29 (3H, s, OMe) and 56.87 (C-3)], a -H₂C-HC=C-C=CH-CH₂- moiety [§ 5.31 (1H, H-7), 5.44 (1H, H-11), 115.27 and 120.49 (each =CH-), and 142.28 and 146.23 (each = C <)], a trisubstituted olefin bond [δ 7.01 (1H, H-24), 129.40 (=C<), and 149.78 (=CH-)] lying between the lactone carbonyl [δ 174.42 (C-26)] and the methine proton attached to the etheral oxygen in the γ -lactone ring [δ 4.98 (1H, H-23)] similar to those of the 4-substituted 2-methyl-2-butenolide ring in compound 6. Detailed analysis of 2D COSY and eims for compound 7 furnished appropriate skeletal information. In the 2D long range ${}^{1}H{}^{-13}C$ nmr spectrum, compound 7 showed cross correlation for signals of Me-18 (with C-12, C-13, C-14, and C-17), Me-19 (with C-1, C-5, C-9, and C-10), Me-21 (with C-17, C-20, and C-22), Me-27 (with C-24, C-25, and C-26), Me-28 (with C-3, C-4, C-5, and C-29), Me-29 (with C-3, C-4, C-5, and C-28) and Me-30 (with C-8, C-13, C-14, and C-15), as well as H-7 (with C-6 and C-8) and H-11 (with C-9 and C-12). The eims of compound 7 exhibited characteristic fragment ion peaks considered to arise from the cleavage of the lanosta-7,9(11)-diene skeleton at m/z 391 [M – Me – MeOH – CO]⁺, 339 [ion **m**], 337 [ion **n**], 327 [ion **o**], 325 [ion **p**], 295 [ion **q**], 285 [ion **r**], and 253 [ion **s**], together with peaks due to cleavage of the side chain moiety containing a 2-methyl-2-butenolide ring

Carbon	Compound						
	2	3	5 ^b	6	6-acetate	7	
C-1	34.20	35.52	31.00	29.82	30.54	30.06	
C-2	34.28	27.94	20.43	25.72	23.23	20.35	
C-3	218.92	79.33	85.51	76.32	78.45	85.67	
C-4	47.01	38.83	37.97	37.40	36.50	37.78	
C-5	52.35	48.62	42.55	43.53	43.33	43.72	
С-6	23.01	23.07	31.30	23.11	23.05	22.99	
С-7	121.64	121.70	72.31	121.64	121.51	120.49	
С-8	148.48	148.59	50.39	148.60	148.53	142.28	
C-9	45.67	48.33	146.43	48.53	48.42	146.23	
C-10	35.80	35.90	39.10	35.69	35.58	37.26	
C-11	20.88	22.93	116.56	22.89	22.85	115.27	
C-12	34.39	35.26	36.83	35.35	35.27	37.65	
C-13	44.15	43.74	45.24	43.73	43.72	43.95	
C- 14	51.93	52.76	46.62	52.88	52.86	50.47	
C-15	33.01	33.32	36.61	33.28	33.24	31.44	
C-16	28.21	28.55	28.58	28.58	28.57	27.88	
C-17	53.47	53.96	50.55	53.98	53.98	51.39	
C-18	22.51	23.66	14.36	23.77	23.78	15.73	
C-19	23.12	24.49	21.89	24.37	24.31	22.71	
C-20	33.49	33.47	33.33	33.47	33.47	33.55	
C-21	18.13	18.41	18.41	18.39	18.41	18.44	
C-22	40.46	40.46	40.73	40.46	40.41	40.60	
C-23	78.96	79.01	78.91	79.02	79.02	78.97	
C-24	149.68	149.66	149.69	149.72	149.72	149.78	
C-25	129.48	129.51	129.52	129.46	129.43	129.40	
C-26	174.44	174.43	174.39	174.44	174.42	174.44	
C-27	10.65	10.65	10.65	10.63	10.63	10.63	
C-28	21.30	16.36	28.22	28.66	28.26	28.27	
C-29	28.00	28.89	22.73	23.41	23.05	23.15	
C-30	27.41	30.45	18.27	30.72	30.80	25.71	
ОМе	— —	—	57.01	_	— —	56.87	
CO_2Me			—		21.30		
CO_2Me		—			170.67		

TABLE 2. ¹³C-nmr Chemical Shifts of Compounds 2, 3, 5, 6, 6-acetate, and 7 in CDCl₃ (TMS = 0).^a

"Assignments were made by 2D 1 H- 1 H COSY, 2D 1 H- 13 C COSY, and 2D long-range 1 H- 13 C COSY experiments.

^bData for this compound are from Tanaka and Matsunaga (6).

analogous to those of compounds 2 and 3 at m/z 139 [ion j], 109 [ion k], and 97 [ion l] (Scheme 2). On the other hand, the cd curve of compound 7 gave a negative Cotton effect (see Experimental) similar to those of compounds 1-6 (1, 3-5), indicative of the same side chain containing the 23*R* configuration of the lactone ring with these compounds. All these data indicated that compound 7 must be 3 α -methoxylanosta-7,9(11),24-trien-26,23*R*-olide. Conclusive evidence for this structure was obtained from the following experiment. Oxidation of abieslactone [1] with selenium dioxide in HOAc furnished the 7,9(11),24-trien-26,23*R*-olide, which was identical in all respects with compound 7.

To the best of our knowledge, compounds 6 and 7 have not yet been described in the literature. Further investigation on the anti-tumor-promoting activity for 6, 7, and their derivatives is now in progress, and results will be reported elsewhere.



SCHEME 2. Mass spectral fragmentation of compound 7.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Melting points were measured on a Yanagimoto micromelting point apparatus and are uncorrected. Optical rotations were taken in CHCl₃ using a Jasco DIP-140 polarimeter. Cd measurements were run in dioxane by employing a Jasco J-500 spectropolarimeter at 25°. Uv spectra were measured in EtOH on a Hitachi 150-20 spectrophotometer, and ir spectra were recorded as KBr discs with a Jasco A-100 ir spectrophotometer. ¹H- and ¹³C-nmr spectra were obtained in CDCl₃ on a Varian XL-300 instrument at 300 MHz and 74.5 MHz, respectively, using TMS as internal standard. Eims were recorded at 70 eV (probe) on a Hitachi M-60 double focusing mass spectrometer. Si gel 60 (Merck, 70–230 mesh) was used for cc. Si gel PF₂₅₄ plates (Merck, 2 mm) were employed for preparative tlc.

ISOLATION OF COMPOUNDS.—Previously, we reported that Si gel cc of the crude crystalline solid (56.5 g) deposited from the syrupy Et_2O extract of the dried stem bark of *A. veitchii* (7.9 kg) led to the isolation of abieslactone [1] and veitchiolide [2] (4). In this stage, resinous products showing closely similar tlc spots have been obtained from the intermediate fractions (fractions 135–165) eluted with CHCl₃ and CHCl₃-EtOAc (20:1). These were combined and condensed to give an amorphous solid (4.3 g), which was subjected to rechromatography on a Si gel (360 g) column. Elution with CHCl₃ afforded compounds 2 (182 mg) and 3 (53 mg), in order of polarity. Successive cc furnished compounds 6 (1.3 g) and 7 (28 mg) from the fractions eluted with the mixed solvent of CHCl₃-EtOAc (30:1) and CHCl₃-EtOAc (20:1), respectively.

COMPOUND 2.—Prisms, mp 245.5–248° (EtOAc), $[\alpha]^{2^3}D + 27.5°$ (*c*=0.55, CHCl₃); *R_f* 0.56 [0.25 mm thick, solvent C₆H₆-CHCl₃-EtOAc (1:1:1)]; hrms [M]⁺ 452.3288 (C₃₀H₄₄O₃ requires 452.3290); ir ν max cm⁻¹ 3070, 3022, 2930, 2852, 1740 (α,β-unsaturated γ-lactone), 1698 (6-membered ring C=O), 1648 (C=C), 1460, 1410 (CH₂CO), 1378, 1360, 1197, 1095, 1040, 933, 878, 818 (-HC=C<); ¹H nmr see Table 1; ¹³C nmr see Table 2; eims *m/z* (rel. int.) [M]⁺ 452 (46), [M – Me]⁺ 437 (100), [M – Me – H₂O]⁺ 419 (13), 315 (11), 314 (12), 313 (11), 299 (20), 271 (18), 243 (20), 139 (9), 97 (37). It was identified by direct comparison (mmp, co-tlc, ir, ¹H-nmr ¹³C nmr, and eims) with an authentic sample of 3-oxo-9β-lanosta-7,24-dien-26,23*R*-olide isolated previously from the stem bark of *A*. *firma*.

COMPOUND **3**.—Needles, mp 239–241° (MeOH), $[\alpha]^{23}D - 58.9^{\circ}$ (c = 0.32, CHCl₃); $R_f = 0.42$ [0.25 mm thick, solvent C₆H₆-CHCl₃-EtOAc (1:1:1)]; hrms m/z [M]⁺ 454.3449 (C₃₀H₄₆O₃ requires 454.3446); ir ν max cm⁻¹ (KBr) 3560 (OH), 3065 (-HC=C<), 2948, 2870, 1760 (shoulder), and 1740

(α,β-unsaturated γ-lactone), 1658 (C=C), 1462, 1378, 1362, 1215, 1106, 1058, 1022, 980, 960, 878, 805 (-HC=C<); ¹H nmr see Table 1; ¹³C nmr see Table 2; eims m/z (rel. int.) [M]⁺ 454 (18), [M – Me]⁺ 439 (35), [M – H₂O]⁺ 436 (4), [M – Me – H₂O]⁺ 421 (100), 315 (7), 314 (20), 299 (22), 273 (4), 245 (4), 227 (11), 139 (14), 97 (86). It was identified by direct comparison (mmp, co-tlc, ir, ¹H nmr, ¹³C nmr, and eims) with an authentic sample of 3β-hydroxy-9β-lanosta-7,24-dien-26,23*R*-olide isolated also from the bark of *A. firma*.

COMPOUND **6**.—Prisms, mp 250–252° (MeOH/CHCl₃), $[\alpha]^{23}D - 80.5°$ (z = 0.55, CHCl₃), $R_f = 0.49$ [0.25 mm thick, solvent C₆H₆-CHCl₃-EtOAc (1:1:1)]; hrms m/z [M]⁺ 454.3443 (C₃₀H₄₆O₃ requires 454.3446); ir ν max cm⁻¹ (KBr) 3480 (OH), 3048 (-HC=C<), 2932, 2850, 1745, and 1728 (shoulder) (α , β -unsaturated γ -lactone), 1645 (C=C), 1458, 1440, 1375, 1365, 1198, 1087, 1015, 968, 925, 862, 810 (-HC=C<); ¹H nmr see Table 1; ¹³C nmr see Table 2; eims m/z (rel. int.) [M]⁺ 454 (20), [M - Me]⁺ 439 (17), [M - H₂O]⁺ 436 (5), [M - Me - H₂O]⁺ 421 (100), [ion **a**] 327 (2), [ion **b**] 315 (11), [ion **c** 314 (32), [ion **d**] 309 (2), [ion **e**] 299 (20), [ion **f**] 273 (7), [ion **g**] 233 (11), 227 (11), [ion **h**] 189 (3), [ion **i**] 187 (32), [ion **j**] 139 (9), [ion **k**] 109 (30), [ion **1**] 97 (55); cd (dioxane) [θ]₂₀₀ - 8100°, [θ]₂₀₈ - 26000° (trough), [θ]₂₁₂ - 22200°, [θ]₂₂₀ - 11500°, [θ]₂₃₀ - 1800°, [θ]₂₄₅ 0°, [θ]₂₅₀ + 200°.

ACETYLATION OF COMPOUND 6.—Compound 6 (100 mg) was dissolved in an equivolume mixture of pyridine and Ac₂O (4 ml). The mixture was kept at room temperature overnight, and subsequent workup afforded 6-acetate (101 mg): m/z [M]⁺ 496.3552 (C₃₂H₄₈O₄ requires 496.3552), mp 204.5–208° (MeOH/CHCl₃); ir ν max (KBr) cm⁻¹ 2965, 2948, 2830, 1740, and 1736 (shoulder) (α ,β-unsaturated γ -lactone), 1722 (OAc), 1465, 1443, 1382, 1370, 1248 (OAc), 1180, 1091, 1048, 1022, 958, 938, 867, 828; ¹H nmr see Table 1; ¹³C nmr see Table 2; eims m/z (rel. int.) [M]⁺ 496.3552 (14), [M – Me]⁺ 481 (16), [M – HOAc]⁺ 436 (11), [M – Me – HOAc]⁺ 421 (100), [ion **a**] 369 (1), [ion **b** – HOAc and ion **f**] 315 (7), [ion **c**] 314 (16), [ion **d**] 309 (2), [ion **e**] 299 (14), [ion **g**] 233 (8), 225 (9), [ion **h**] 189 (8), [ion **i**] 187 (20), [ion **j**] 139 (3), [ion **k**] 109 (10), [ion **l**] 97 (2).

OXIDATION OF COMPOUND **6** WITH CrO₃ IN PYRIDINE.—A solution of CrO₃ (50 mg) in pyridine (3 ml) containing 1 drop of H₂O was gradually added to a solution of compound **6** (50 mg) in pyridine (5 ml) at 10°, while stirring, and the mixture was kept at 20° for 5 h. Then, 2 drops of 10% NaHSO₃ solution was added to destroy excess CrO₃. Removal of the pyridine in vacuo gave a residue which was dissolved in Et₂O (30 ml), washed with 5% HCl and H₂O, and dried over Na₂SO₄. Removal of solvent yielded a solid which was purified by preparative tlc [C₆H₆-CHCl₃-EtOAc (1:1:1)] to give the ketone, mp 245–247° (EtOAc), 46.5 mg. The ketone was identified by direct comparison (mmp, co-tlc, ir, ¹H nmr, ¹³C nmr, and eims) with an authentic sample of 3-0xo-9β-lanosta-7,24-dien-26,23*R*-olide [**2**] isolated from the stem bark of both *A. firma* and *A. veitchii*.

COMPOUND 7.—Prisms, mp 208–209.5° (MeOH/CHCl₃), $[\alpha]^{23}$ D + 1.4° (r = 0.65, CHCl₃); hrms m/z [M]⁺ 466.3451 (C₃₁H₄₆O₃ requires 466.3447); uv λ max (ErOH) nm 236, 243.5, 251.5 (log ϵ 4.26, 4.32, 4.15) (heteroannular diene); ir ν max (KBr) cm⁻¹ 2935, 2870, 1758 (shoulder), 1740 (α , β -unsaturated γ -lactone), 1658 (C=C), 1445, 1420, 1377, 1360, 1205, 1100, 1063, 940, 897, 815 (-HC=C<); ¹H nmr see Table 1; ¹³C nmr see Table 2; eims m/z (rel. int.) [M]⁺ 466 (100), [M – Me]⁺ 451 (8), [M – MeOH]⁺ 434 (14), [M – Me – CO]⁺ 423 (5), [M – Me – MeOH]⁺ 419 (43), [M – Me – MeOH]⁺ 4391 (6), 383 (4), 365 (6), 351 (3), [ion m] 339 (6), [ion n] 337 (11), [ion o] 327 (4), [ion p] 325 (5), [ion i] 187 (12), 171 (16), [ion j] 139 (3), [ion k] 109 (8), [ion l] 97 (20); cd (dioxane) [θ]₂₀₀ – 6700°, [θ]₂₀₈ – 19800°, [θ₂₁₂ – 19900° (trough), [θ]₂₃₀ – 7600°, [θ]₂₄₀ – 4900°, [θ]₂₆₁ 0°, [θ]₂₇₀ + 450°.

SYNTHESIS OF COMPOUND 7 FROM ABJESLACTONE [1].—A solution of freshly sublimed selenium dioxide (20 mg) in 80% HOAc (5 ml) was gradually added to a solution of abieslactone [1] (40 mg) in HOAc (10 ml), and the mixture was refluxed for 1 h. After cooling, the reaction mixture was diluted with H_2O (40 ml), the resulting precipitate was extracted with Et_2O (50 ml), and the Et_2O layer was neutralized and dried with Na_2SO_4 . Removal of the Et_2O afforded a residue (38 mg), which was purified by preparative tlc [C_6H_6 -CHCl_3-EtOAc (1:1:1)] to give 3α -methoxylanosta-7,9(11),24-trien-26,23*R*-olide (30.5 mg), mp 206–208.5° (MeOH/CHCl_3), [α]²³D + 1.4° (c = 1.02, CHCl_3), m/z [M]⁺ 466.3445, uv λ max (EtOH) nm 236, 243.5, 251.5 (heteroannular diene). This compound was identified by direct comparison (mmp, co-tlc, uv, ir, ¹H nmr, ¹³C nmr, and eims) with compound 7.

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LITERATURE CITED

- 1. Y. Hayashi, in: "Plants of the World, Asahi Encyclopedia." Ed. by S. Kitamura, M. Honda, and T. Satoh, Asahi Shinbun-sha, Tokyo, 1977, Vol. 106, p. 2512.
- 2. S. Uyeo, J. Okada, S. Matsunaga, and J.W. Rowe, Tetrahedron, 24, 2859 (1968).
- 3. F.H. Allen, N.W. Isaacs, O. Kennard, and W.D.S. Motherwell, J. Chem. Soc., Perkin Trans. 2, 498 (1973).
- 4. R. Tanaka, A. Inoshiri, M. Yoneda, T. Ishida, A. Numada, and S. Matsunaga, *Phytochemistry*, 29, 3263 (1990).
- 5. R. Tanaka and S. Matsunaga, Phytochemistry, 30, 1983 (1991).
- 6. R. Tanaka and S. Matsunaga, Phytochemistry, 29, 3267 (1990).
- 7. J.-C. Muller and G. Ourisson, Phytochemistry, 13, 1615 (1974).
- 8. W. Steglich, M. Klaar, L. Zechlin, and H.J. Hecht, Angew. Chem., 91, 751 (1979).
- 9. J.P. Kutney, D.S. Grierson, G.D. Knowles, N.D. Westcott, and I.H. Rogers, Tertrahedron, 29, 13 (1973).
- 10. S. Hasegawa, T. Miura, N. Kaneko, Y. Hirose, and Y. Iitaka, Tetrahedron, 43, 1775 (1987).
- 11. S. Hasegawa, N. Kaneko, and Y. Hirose, Phytochemistry, 26, 1095 (1987).
- 12. V.A. Raldugin, Yu. V. Gatilov, T.V. Ryabalova, and Ya. V. Rashkes, *Khim. Prir. Soedin.*, 688 (1986), and references cited therein.
- 13. V.A. Raldugin, S.A. Shevtsov, N.I. Yaroshenko, Yu. V. Gatilov, I. Yu. Bagryanskaya, L.I. Demenkova, and V.A. Pentegova, *Kbim. Prir. Soedin.*, 824 (1987).
- 14. J. Takayasu, R. Tanaka, S. Matsunaga, H. Ueyama, H. Tokuda, T. Hasegawa, A. Nishino, H. Nishino, and A. Iwashima, *Cancer Lett.*, **53**, 141 (1990).

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